

An assessment of the evidence on antiseptics: a consensus paper on their use in wound care

Antiseptics are indicated for wounds with localised infection, but there is little robust evidence to guide selection. This consensus paper explores the clinical and research literature to make recommendations for their use in day-to-day practice

wound antiseptics; povidone iodine; octenidine dihydrochloride; polyhexanide; topical antibiotics

To date 'evidence-based' recommendations for the use of antiseptics on acute and chronic wounds are based on various levels of knowledge (**okay?**), including clinical experience. No randomised controlled double-blind clinical studies with recognised parameters and comparative procedures for applying antiseptics have been conducted, although first steps are being taken.¹

In the light of the generally unsatisfactory data on wound antiseptics, this paper set out to investigate active agents that are considered to:

- Have a reliable broad-spectrum effect
- Have a rapid onset of effect^{2,3}
- Be efficacious under organic stress (that is, are active when applied in an organ system as opposed to single cells)^{2,3}
- Promote wound healing
- Have adequate cell and tissue tolerance (that is, are not toxic)
- Prevent allergy, anaphylaxis, reabsorption and development of resistance.

Author, could you add a sentence or two on this being a consensus document, how the consensus committee was formed, and how you came to the consensus. Thanks Conclusions are based on evidence gained from both *in vitro* studies and clinical practice.

Indications for use

Accurate indications for the use of antiseptics are vital to avoid inhibition of healing or wound damage.⁴ Contamination or colonisation of wounds, unless by methicillin-resistant *Staphylococcus aureus* (MRSA), is relatively common and generally does not affect the healing process.⁵ However, patients with burns are particularly at risk from contamination due to the extensive wound surface and the presence of non-vital tissue and exudate in the wound bed.⁶

Wound infection can be classed as primary or secondary. Trauma wounds, particularly bites, traffic

injuries and stab wounds, have the potential for primary infection — surface microorganisms can migrate into the deeper tissues — so antiseptic prophylaxis is required. Infection developing in an existing wound is secondary.

Generally, localised infection should be treated with antiseptics, systemic infection requires antibiotics, and life-threatening infections such as streptococci in acute necrotising fasciitis need appropriate interventions **such as?**

Antiseptics: short-term use

The primary aim is to eliminate microorganisms in the wound by surgical debridement, where appropriate, wound cleansing and coverage.^{2,3,7,8}

Contaminated injuries with good wound access and intact tissue perfusion require one just application of wound antiseptic. Clinically infected wounds should be cleansed with an antiseptic until the infection is eliminated.⁹

Povidone-iodine

Povidone-iodine is effective against Gram-positive and Gram-negative bacteria, fungi and protozoa and, with a longer exposure time, spores¹⁰ and a range of viruses.¹¹⁻¹³ Like octenidine (Octinisept, Schülke and Mayr), it has a rapid antimicrobial effect — within 30 seconds without organic stress *in vitro*.¹⁴⁻²⁰ **(TC to check if is available in the UK)**

Studies have demonstrated the activity and efficacy of povidone-iodine **combined with (correct?) octenidine/phenoxyethanol (again, not in BNF. UK equivalent?).**^{20,21} Both iodine and octenidine are effective against vegetative bacteria (bacteria in a passive, unproductive state), although octenidine is ineffective against spores and protozoa.²²⁻²⁵

Iodine is better tolerated by tissue than a combination of octenidine/phenoxyethanol or preparations containing chlorhexidine, but less well tolerated than polyhexanide and taurolidine **(again, not in BNF. UK equivalent?).**²⁶⁻²⁹ **(But are polyhexanide and taurolidine less effective than**

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References

1 Reimer, K., Vogt, P.M., Brögmann, B. et al. An innovative topical drug formulation for wound healing and infection treatment: *in vitro* and *in vivo* investigations of a povidone iodine liposome hydrogel *Dermatology* 2000; 201: 235-241.

2 Kramer, A., Heeg, P., Harke, H.P. et al. Wundantiseptik. In: *Klinische Antiseptik* 1993; 163-191

3 Kramer, A., Wendt, M., Werner, H.P. Möglichkeiten und Perspektiven der klinischen Antiseptik mhp, Wiesbaden 1995

4 Kramer, A. Replik zum Artikel "Aktuelles Präparatespektrum und Anwendungs-emphelungen für die Wundantiseptik" im *Hygiene Monitor* 8/99 *Hygiene Monitor Jahrgang* 5: 12/99

5 Zastrow, K.D., Kramer, A. Recommendations for Isolation and Antiseptic Sanitation of Patients with MRSA Colonisation or Infection In: Kramer A, Behrens-Baumann W (eds): *Antiseptic Prophylaxis and Therapy of Ocular Infections* Karger, Basel, 2001.

6 Tompkins, R.G., Burke, J.F. *Infections of Burn Wounds* In: Bennet, J.V., Brachman, P.S. (eds) *Hospital Infections* (3th edn). Little Brown, 1992.

7 Baharestani, M. The clinical relevance of debridement In: Baharestani, M., Goltrup, F., Holstein, P., Vanscheidt, W. (eds) *The Clinical Relevance of Debridement*. Springer, 1999.

8 Dräger, E., Winter, H. Surgical debridement versus enzymatic debridement. In: Baharestani, M., Goltrup, F., Holstein, P., Vanscheidt, W. (eds) *The clinical relevance of debridement*. Springer, Berlin Heidelberg 1999

9 Kramer, A., Bergemann, R., Bruck, J. et al. Indikationen und Auswahlkriterien für lokale Wundantiseptika und Wundaufgaben im Rahmen der chirurgischen Wundbehandlung *Losebattsammlung Hygiene in Krankenhaus und Praxis* 1-20.5, ecomed Landsberg, 1-16, 1999.

10 Gershenfeld, L. Povidone-iodine as a

Box 1. Indications for application of povidone-iodine, as specified in the manufacturer's instructions and instructions for use

Single application

Antisepsis of the intact external skin

Antisepsis of mucus membranes — for example, before surgical interventions, biopsies, injections, punctures and catheterisation of the bladder

Repeated, temporarily limited application

Antiseptic wound management — for example, pressure ulcers, leg ulcers, burns

Infected and superinfected dermatoses

Hygienic and surgical hand disinfection

iodine etc?)

As an active component, therefore, iodine is the agent of choice for topical management of infected wounds or colonised acute trauma wounds.^{13-16,18} It can also be used for rinsing deep wounds and body cavities, such as pleura, in a 1:10 solution.³⁰⁻³² It is also suitable for pre- and postoperative antiseptic application and is the first choice for pre-operative use in eye surgery.^{17,33-37}

A combination of 39 w/w% each of ethanol and (okay?) 2-propanol with povidone-iodine is the first choice of antiseptic in stab wounds or lacerations in patients with HIV, hepatitis B or hepatitis C.^{38,39}

Intra-articular 0.5% povidone-iodine was well tolerated in rabbits.³⁹ This was later confirmed *in vitro* on adult bovine cartilage.⁴⁰ Studies have demonstrated that tissue compatibility is significantly improved, with no loss of effectiveness, when povidone-iodine is mixed into a liposomal preparation. *In vitro*, even enhanced cell proliferation was observed.⁴⁰

Indications for the use of povidone-iodine are given in Box 1.

In animal experiments, iodophors do not trigger allergic reactions. In humans, this happens rarely.²² However, the following are contraindicated:

- Hyperthyroidism
- Dermatitis herpetiformis
- Dühring's disease (a polyaetiological syndrome involving focal infections, malign tumors and allergic processes)
- Iodine hypersensitivity
- Radio-iodine therapy.

Iodine is not advised following skin grafting¹ or peritoneal lavage due to the increased risk of tissue intolerance (povidone-iodine can deposit in the liver, and/or adhesiolysis [please define briefly] or a shift in the acid-base balance may occur).^{23,41}

Finally, clinicians must remember that, depend-

Box 2. Indications for octenidine in combination with phenoxyethanol according to the manufacturers' instructions for application

For repeated, temporally limited antisepsis of mucous membranes and adjacent skin before diagnostic interventions and surgical procedures in the anal/genital area, in the oral cavity, and for temporally limited supporting therapy of interdigital mycoses, as well as adjuvant antiseptic wound management

ing on the components and concentration of the active agent in the product, the proportion of freely available iodine can vary, influencing its effect.

Octenidine dihydrochloride

Octenidine dihydrochloride, a surface-active agent, is used either in combination with 2% phenoxyethanol or as the sole active agent (in cosmetics). Its antimicrobial activity extends to Gram-positive and Gram-negative bacteria, fungi and certain viruses, but it is ineffective against spores and protozoa.⁴²⁻⁴⁴

In contrast to iodophors, the exposure time of a 1:1 dilution of an octenidine/phenoxyethanol-based antiseptic without organic stress varies from 30 seconds to over five minutes,⁴³ depending on the MRSA strain (author, was this just used on MRSA strains?). Against other vegetative pathogens, the full effect unfolds only after five minutes.²⁸ There is no evidence of carcinogenic, mutagenic, teratogenic, embryotoxic or fertility-impairing activity.²²

When applied to wounds there is no observed reabsorption.²² Like povidone-iodine-based antiseptics, dermal application in experimental animals showed no indication of systemic side-effects or neurotoxic reactions.⁴⁴

Cell and tissue toxicity of the commercially available combination of octenidine and phenoxyethanol (Octenisept, Schülke and Mayr) is similar to that of polihexanide. This contradicts empirical clinical reports of successful antisepsis of abrasions, bites and cuts.⁴⁵ Surprisingly, in cell cultures, undiluted octenidine seemed less cytotoxic than a diluted octenidine solution. **reference?** This could explain the discrepancy between current *in vitro* findings — obtained with diluted octenidine solutions — and clinical observations.

Occlusive applications with products containing octenidine or povidone-iodine — for example, in combination with bandages or special dressings — are only to be used if recommended by the manufacturer.

Active agents for long-term use

Here, the objective is to interrupt the vicious circle of colonisation → infection → recolonisation → reinfection → delayed wound healing and to eliminate local or systemic factors that delay healing, with the aim of establishing an optimal wound environment.

Table 1. Further treatment options for colonised or infected wounds

Means/method	Effect per single application/side-effects		
	Minutes to hours	Hours to 1 day	Hours to days
(1–7 days)			
Silver dressings ^{89,90}			(depending on the product – different duration of activity)
Special measures for local infection treatment			
VAC therapy with polyurethane foam ⁷¹		(daily change of dressing advised by the manufacturer in case of wound infection)	
VAC therapy with polyurethane foam ⁷¹			(may be combined with lavage)
Fly larvae ^{70,92–96} (<i>Lucilia sericata</i>)			(1–4 days)

Polihexanide

Depending on the pathogen and concentration of the agent, the microbicidal activity of polihexanide is slower than iodophores and octenidine (0.04% *in vitro* within 5–20 minutes). It is not effective against viruses and spores, but is effective against *Acanthamoeba keratitis*.^{22,19,46–50} Good tissue compatibility — caused by its activity against acid lipids of *in?* bacterial cell membranes, its minor effect on the neutral lipids of human cell membranes,⁵¹ its clinical effectiveness and its ability to support the formation of granulation tissue — makes polihexanide the first choice for non-healing chronic and/or refractory wounds such as second-degree burns and lavage.^{22,29,52–59}

According to current knowledge, it appears that, due to its molecular size, polihexanide is not reabsorbed and therefore remains effective only at the site of application.

Initial experiments by Brunner et al. 2003 (personal communication) suggested that polihexanide is compatible with products such alginates and hydrofibres. Due to tissue compatibility and the absence of irritation, application under semi-occlusive and occlusive dressings is possible.⁴⁶

In Germany and Austria polyhexanide is available as a pharmaceutical raw material for the manufacturing of pharmacy-prepared solutions for wound antiseptics. In Switzerland it is registered as a concentrate and pharmacy-prepared solution. In addition, a wound rinse containing undecylenamide-propyl-betaine (**author, please check this is correct. I couldn't find it on the Internet or in the British National Formulary**) as a surface active substance and polyhexanide (combination preparation) as a 'preservative' is available **in which country?** for wound cleansing, moistening and flushing of germs (Sanalind, Paul Hartmann).

Contraindications for polihexanide preparations include:

- Allergies to the active agent and/or ingredients of the applied product
- Application on the hyaline cartilage, central nervous system, the middle and inner ear and inner eye
- First four months of pregnancy.²²

Polyhexanide may not be used in combination with anionic tensides (**author, tensides is not in any dictionary, please check it is correct and define what it means**) or other wound cleansing soaps, ointments, oils and enzymes etc.⁶⁰

Taurolidine

The active agent taurolidine (**again, not in BNF. UK equivalent?**) has two specific characteristics.^{61–65} Due to the slow formaldehyde release⁶¹ *in vitro*, the necessary bactericidal action (reduction factor over 5lg-steps - **author, please explain briefly what g-steps are**) only unfolds after 6–24 hours.⁴⁸ It remains effective in the presence of proteins and blood. (**Author, are these the two characteristics? If so, why are they referenced with refs. 62–65. Please clarify**)

Further active agents and methods

Supplemental treatment is needed for problem patients (Table 1). Biosurgery is significantly superior to conventional treatment procedures and usually well accepted by patients,^{66–78} although some have occasionally reported pain during application.^{6,71,76}

Larvae aid effective debridement and reduce the number of bacteria by up to 5 lg-steps *in vitro*.⁷¹ Indeed, MRSA wound infections have been successfully treated with larvae,^{72,73} and preparations containing their haemolymphatic and alimentary secretions have been shown to stimulate fibrob-

sporicide Am J Pharm 1962; 134: 78–81.

11 Esanu, V., Profeta, A. Antiviral Antiseptics In: Kramer A, Krasilnikow AP, Weuffen W, Berencsi G, Gröschel D, Kemter BP (Hrsg) Handbuch der Antiseptik, Bd. II/3, Antibakterielle, antifungielle und antivirale Antiseptik –ausgewählte Wirkstoffe (Hrsg. Kramer A, Weuffen W, Krasilnikow AP, Gröschel D, Bulka E, Rehn D), Fischer, 1987.

12 Wutzler, P., Sauerbrei, A., Klöcking, R. et al. Virucidal and chlamydicidal activities of povidone-iodine liposome complex Ophthalmic Res 2000; 32: 118–125.

13 Daróczy, J. Antiseptic efficacy of local disinfecting povidone-iodine (Betadine®) therapy in chronic wounds of lymphedematous patients Dermatology 2002; 204: Supp 1, 75–78.

14 Görtz, G., Reimer, K., Neef, H. Entwicklung, Eigenschaften und Bedeutung von PVP-Iod In: Topische Infektionstherapie und Prophylaxe 1996; 3–7.

15 Michel, D., Zäch, G.A., von Arx, P., Geng, V. Wachstumshemmende Wirksamkeit von Antiseptika im Suspensionstest *in vitro* auf Methicillin-resistente Staphylococcus aureus-Stämme (MRSA), Pseudomonas aeruginosa und Escherichia coli In: Topische Infektionstherapie und Prophylaxe: 9–12, 1996.

16 Kramer, A., Rudolph, P., Pitten, F.A. et al. Glück U Antiseptika im Kampf mit den Keimen Pharm Ztg 2000; 145: 2, 11–19.

17 Kramer, A., Behrens-Baumann, W. Prophylactic use of topical anti-infectives in ophthalmology Ophthalmologica 1997; 211: Supp 1: 68–76.

18 Mlangeni, D., Daschner, F. Povidone-iodine: Evaluation of povidone-iodine as an antiseptic. Antiinfective Drugs Chemother 1995; 13: 3, 161–167.

19 Werner, H.P. Die mikrobizide Wirksamkeit ausgewählter Antiseptika Hyg Med 1992; 17: 2, 51–59.

20 Pitten, F.A., Werner, H.P., Kramer, A. A standardized test to assess the impact of different organic challenges on the antimicrobial activity of antiseptics J

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Table 2. Characteristics of obsolete or dispensable active agents for wound antiseptics

Active	Advantage	Disadvantage	Suitable for wound antiseptics
8-Chinololinol	None	insufficiently effective, mutagenic, neurotoxic, allergenic, in animal studies carcinogenic	Dispensable (of minor importance or not essential)
Chloramin T	None	Insufficiently effective, inactivation by blood, allergenic, cytotoxic	Dispensable
Chlorhexidine	Remnant	Gap in efficacy spectrum, cytotoxic, mutagenic, reversible pre-malignant changes in the mouth of rats, anaphylaxis, neurotoxic, resorption?	Dispensable, not for application in the peritoneal area
Ethanol	10% enhanced wound healing in vitro ^{4,9}	70% solution causes stinging	10% solution in combination with other antiseptics useful, 70-80 % solution where alternatives unavailable may be used as a stand alone
Ethacridine lactate	None	Allergenic, delays wound healing, in vitro mutagenic, more toxic than modern antiseptics (sc LD50 about 1/20 of PVP-I), insufficiently effective, resistance development, not stable under the influence of light	Obsolete
Dyes	None	Insufficiently effective, topical sensitisation, possible systemic risks (authors, correct?)	Obsolete
Nitrofurantoin	None	Insufficiently effective, mutagenic, allergenic, induced benign tumors, resorption in wounds, resistance development possible	Dispensable
Organic mercury compounds	None	Pathogen-dependent, sometimes ineffective, systemic side effects, sensitizing, environmental impact	Obsolete
Quats	None	Insufficiently effective, cytotoxic, resorptive risks, resistance development	Dispensable
SSD (silver sulphadiazine)	Temporarily comfortable, cooling	Insufficient microbicidal activity in vitro, resistance development, cytotoxic, systemic risks, allergenic, formation of disturbing protein-wound exudate complexes (scab)	Dispensable
Hydrogen peroxide 3%	Cleansing intact skin from e.g. blood particles via O ₂ formation	Insufficiently effective, inactivated by blood, cytotoxic	Dispensable

lasts, which reached 12% of the amount of the stimulation induced by epidermal growth factor (EGF).⁷⁸ When compared with hydrogels, the cost of larvae therapy was found to be significantly lower due to accelerated wound closure, lower material costs and the reduced need for antibiotics.⁶

For different reasons, such as effectiveness, tolerance/toxicity, the antiseptics listed in Table 2 are not suitable for general use, or are reserved for special situations **such as?**^{2,3,9,17,19,22,29,79-83}

Silver sulphadiazine

Silver sulphadiazine, a complex of silver and sulphadiazine (a sulphone amide), is used in the treatment of burns before surgical necrotomy. As its benefit:risk ratio is being viewed ever more criti-

cally, a more detailed assessment seems appropriate.

It can be assumed that, with the use of microbostatically active agents such as silver sulphadiazine, an effect can only be expected if the bacterial burden is low (less than 10⁵ CFU (**please write in full**)/g tissue).⁷⁹ The cytotoxicity⁸⁰⁻⁸⁴ of this active agent could well be the cause of the delayed epidermal regeneration, which occurs alongside transient signs of a reaction similar to dermatitis, with spongiolysis, parakeratosis and pseudocarcinomatosis.⁷⁹

When applied to burn injuries, silver concentrations in the blood of up to 440µg/l and in urine of up to 12µg/l have been measured, which may become toxicologically and **allergologically** (**author, is this a proper word?**) relevant.⁸³ It is advisable, therefore, to monitor silver absorption in ►

PICTURE CREDIT

blood and/or urine.

In patients with sulphone amide hypersensitivity and renal insufficiency, the use of silver sulphadiazine is strictly contraindicated and the possibility of developing resistance to silver ions, a cross-resistance to systemically administered sulphonamides, should be considered.⁸⁴

Applying silver nitrate to chronic wounds before skin grafting has induced deep necrosis and surface oedema of the corium and/or fatty tissue, as well as fibrin deposits. **reference 85?** In superficial fibrin, minor infiltration by cylindrical cells and granulocytes was observed **reference 85?** In the deep vessels endothelial cells swelled, and there was leucostasis and a leucocytoclastic penetration of the vessel walls, which could be an expression of a toxic substance reaction. The skin layer directly on the surface consisted of virtually only a necrotic zone with granulocyte infiltration.⁸⁵

Obsolete or dispensable active agents

This includes all substances and combinations of substances that — due to their uncertain effectiveness, critical cytotoxicity, irritation and allergy potential, pain induction, development of resistance and/or absorptive risks — are not or are no longer recommendable for application, or for which proof of efficacy is lacking.

Wounds requiring immediate surgical intervention, such as necrotising fasciitis and deep dermal burns, should not be given antiseptics as a first-line treatment. Necrotising fasciitis is a rare, rapidly progressive, soft-tissue infection characterised by extensive necrosis of the skin and subcutaneous tissue. For both wound types, use of an antiseptic may be considered after surgical intervention.⁸⁶

Topical antibiotics

These products, which include neomycin, kanamycin (**again, TC to check in BNF. UK equivalent?**) and mupirocin, can only be applied topically due to their lack of absorption and/or systemic toxicity. However, their use is opposed because of their:

- Narrow spectrum of effectivity
- Inadequate — essentially only microbiostatic — efficacy⁸⁷
- Potential for resistance and cross-resistance
- Insufficient or no activity against multiresistant pathogens such as MRSA
- Lack of remnant efficacy — for example, due to local metabolism
- Insufficient concentration at the site where the effect is required
- Cytotoxic potential in long-term use, often already in short-term use⁴¹
- Pronounced allergy potential.⁸⁸

Antimicrobial chemotherapeutics

When treating infected wounds, as in prophylaxis, clinicians must determine whether the infection can be controlled with topical agents or if adjuvant systemic antimicrobial agents are necessary. The following should be considered:

- Antiseptics with a microbicidal effect such as iodophors, octenidine, polyhexanide are more effective than microbiostatic topical antibiotics. For example, a number of antibiotics have failed to decontaminate nasal MRSA,⁸⁷ whereas iodophors have succeeded.⁵²
- Used and selected correctly, antiseptics are less cytotoxic than antibiotics⁴¹
- Topical application ensures an antiseptically effective concentration in tissue without producing an antimicrobially effective concentration in the rest of organism, thereby reducing the risk of systemic side-effects

Conclusion

In contrast to antibiotics, wound antiseptics are available which have no allergenic risks due to the structure of their active agents.

Although the above recommendations will help support decision-making, they do not represent all of the scientific data relevant for deciding which antiseptic product is indicated for which type of wound.

- Hosp Inf, im Druck 2003
- 21 Müller, G., Kramer, A. In vitro action of combinations of selected antimicrobial agents and adult bovine articular cartilage (sesamoid bone) Chem-Biol Interactions 2003; 145: 331-336.
- 22 Kramer, A. Antiseptika und Händedesinfektionsmittel. In: Korting HC, Sterry W (Hrsg) Therapeutische Verfahren in der Dermatologie Blackwell Wissenschaft Berlin, 2001.
- 23 Hierholzer, G., Görtz, G. PVP-Jod in der operativen Medizin. Grundlagen, klinische Anwendung und Ergebnisse In: PVP-Jod in der operativen Medizin: 280; 1984
- 24 König, B., König, W., Reimer, K. Jod - die Stellung eines alten Desinfektionsmittels in der modernen Infektiologie Dtsch Med Wochenschr 1997; 122: 5, 141.
- 25 Görtz, G. PVP-Jod zur Prophylaxe und Therapie von Infektionen in der Allgemein- und Viszeralchirurgie In: Topische Infektionstherapie und Prophylaxe 1996; 61-68.
- 26 Kramer, A., Adrian, V., Adam, C. Vergleich der Toxizität von Lavasept und ausgewählten Antiseptika Hyg Med 1993; 18: 1:9-16.
- 27 Kramer, A., Adrian, V. Antiseptika als Alternative zu systemischen Antinfektiva mit Ergebnissen zur Gewebeverträglichkeit im Explantationstest als einem weiterentwickelten In-vitro-Prüfmodell. In: Hierholzer G, Reimer K, Weissenbacher ER (Hrsg) Topische Infektionstherapie und Prophylaxe, Thieme, 1996.
- 28 Kramer, A., Adrian, V., Rudolph, P., Kühl, H. In-Vitro-Prüfung der Verträglichkeit ausgewählter antiseptischer Wirkstoffe bzw. Präparate In: Kramer A, Wendt M, Werner HP (Hrsg) Möglichkeiten und Perspektiven der Klinischen Antiseptik. mhp, Wiesbaden, 1995.
- 29 Kramer, A., Adrian, V., Rudolph, P. et al. Explantationstest mit Haut und Peritoneum der neonatalen Ratte als Voraussagetest zur Verträglichkeit lokaler

Antinfektiva für Wunden und Körperhöhlen Chirurg 1998; 69: 8, 840-845.

30 Neef, H., Meyer, M., Fischbeck, O. Erfahrungen in der therapeutischen und prophylaktischen Anwendung von PVP-Iod im Thoraxbereich In: Topische Infektionstherapie und Prophylaxe 1996; 51-56.

31 Stobernack, A., Achatz, R. Behandlungskonzepte zur Infektionsvermeidung und Therapie infektiöser Komplikationen in der Thorax- und Gefäßchirurgie In: Topische Infektionstherapie und Prophylaxe 1996; 57-59.

32 European Tissue Repair Society (1997) Iodine revisited. ETRS Bull 4 (1): 2-5

33 Görtz, G. Intraoperative Spülung mit antiseptischen Lösungen In: Infektionsverhütung in der Chirurgie 1991; 291-303.

34 König, B., Reimer, K., Fleischer, W., König, W. Effects of Betaisodona on parameters of host defence Dermatology 1997; 195: Supp 2: 42-48.

35 Behrens-Baumann, W., Kramer, A. Pre-, Intra- and Postoperative Antisepsis in Eye Surgery In: Kramer A, Behrens-Baumann W (eds) Antiseptic Prophylaxis and Therapy in Ocular Infections. Karger, 2002.

36 Hara, J., Yasuda, F., Higashitsumi, M. Preoperative antiseptics of the conjunctival sac in cataract surgery. Ophthalmologica 1997; 211: (Suppl): 62-67.

37 Binder, C., de Kaspar, H.M., Engelbert, M. et al. Bakterielle Keimbesiedelung der Konjunktiva mit Propionibacterium acnes vor und nach Polyvidon-Jod-Applikation vor intraokularen Eingriffen Ophthalmologie 1989; 95: 438-441.

38 Binder, C., de Kaspar, H.M., Klaus, V., Kampik, A. Präoperative Infektionsprophylaxe mit 1%iger Polyvidon-Jod-Lösung am Beispiel von konjunktivalen Staphylokokken Ophthalmologie 1999; 96: 666-673

39 German-Austrian recommendation for post-exposure prophylaxis of HIV infection (1999): AWMF-online Leitlinie www.AWMF/II/aids004e.ht

m

40 Ganzer, D., Völker, L., Follak, N. Reaktion des hyalinen Gelenknorpels und der Synovialis auf eine intraartikuläre Instillation von verschiedenen Antinfektiva Arthroskopie 2001; 14: 31-44.

41 Kramer, A., Below, H., Behrens-Baumann, W. et al. New aspects of the tolerance of the antiseptic povidone-iodine in different ex vivo models Dermatology 2002; 204, supp 1: 86-91.

42 Harke, H.P., Streek, M. Octenidine - ein neuer antimikrobieller Wirkstoff Hyg Med 1989; 14: 372-374.

43 Harke, H.P. (1997): Moderne Schleimhautantiseptika - Octenidin-dihydrochlorid Vorgestellt auf dem 2. Ulmer Hygiene-Symposium

44 Kramer, A., Mersch-Sundermann, V., Gerdes, H. et al. Toxikologische Bewertung für die Händedesinfektion relevanter antimikrobieller Wirkstoffe. In: Kampf G (Hrsg) Hände-Hygiene im Gesundheitswesen, Springer, Berlin, 2003.

45 Schülke & Mayr GmbH (1997): Adjuvante und unterstützende Wundversorgung mit Octenisept® Firmenschrift 1086/II/5.0/5/97/vDuN

46 Bruck, J.C., Koch, S., Kramer, A. Klinische und histologische Untersuchungen zur Wirksamkeit von Lavasept auf granulierenden bzw. Epithelisierenden Wunden Hyg Med 2000; Suppl. 1: 46.

47 Falanga, V. Classifications for wound bed preparation and stimulation of chronic wounds Wound Repair Regen 2000; 8: 347-350.

48 Skripitz, R., Werner, H.P. Bakterizide Langzeitwirkung ausgewählter Antiseptika Hyg Med 1994; 19: 4: 199-204.

49 Kramer, A., Rudolph, P. Efficacy and Tolerance of Selected Antiseptic Substances in Respect of Suitability for Use on the Eye In: Kramer, A., Behrens-Baumann, W. (eds) Antiseptic Prophylaxis and Therapy in Ocular Infections. Karger, Basel, 2002.

50 Berg, A. Einfluß der peritonealen Spülung mit dem Antinfektivum Polihexanid/Lavasept, auf

die experimentell induzierte Peritonitis beim Meerschweinchen Diss Med Fak Univ Greifswald, 2000.

51 Ikeda, T., Tazuki, S., Watanabe, M. Interaction of biologically active molecules with phospholipid membranes 1. Fluorescence depolarization studies on the effect of polymeric biocides bearing biguanide groups in the main chain Biochem Biophys Acta 1983; 735: 380-386.

52 Kallenberger, A., Kallenberger, C., Willenegger, H. Experimentelle Untersuchungen zur Gewebeerträglichkeit von Antiseptika Hyg Med 1991; 16: 10: 383-395.

53 Kramer, A., Glück, U., Heeg, P., Werner, H.P. Antiseptik. In: Krankenhaus- und Praxishygiene: 252-268

54 Sellmer, W. Lokaltheraeutika, speziell Antiseptika, in der Behandlung chronischer Wunden - eine aktuelle. Bewertung Med Praxis 2001; 2: 20-30.

55 Roth, B., Müller, J., Willenegger, H. Intraoperative Wundspülung mit einem neuartigen lokalen Antiseptikum Helv Chir Acta 1985; 52: 61-65.

56 Willenegger, H. Lokale Antiseptika in der Chirurgie - eine Wiedergeburt? Unfallchir 1995; 20: 94-110.

57 Willenegger, H. Klinische Erfahrungen mit einem neuen Antinfektivum

Arbeitstagungen Liesetal 1992 und 1993. Hyg Med 1994; 19: 4, 227-233.

58 Kramer, A., Willenegger, H. Editorial. Perioperative Antibiotikaphylaxe - dominierende Möglichkeit zu Infektionsprophylaxe bei chirurgischen Eingriffen? Hyg Med 1994; 19: 4, 180-182.

59 Schmit-Neuerburg, K.P., Bettag, Ch., Schlickewei, W. et al. Wirksamkeit eines neuartigen Antisepticum in der Behandlung kontaminierter Weichteilwunden Chirurg 2001; 72: 61-71.

60 Weuffen, W., Kramer, A., Paetzelt, H., Lüdde, K.H. Biologische Bedeutung von Thiocyanat und Schlußfolgerungen für die lokale Infektabwehr In:

Weuffen W, Berencsi G, Gröschel D, Kemter BP, Kramer A, Krasnikow AP, Handbuch der Antiseptik. Bd I/4, S 218-257

61 Reding, R., Pfirrmann, R.W. Taurolidine peritoneal lavage as prophylaxis against infection after elective colorectal surgery [letter; comment] Br J Surg 1995; 82: 569.

62 Billing, A., Frohlich, D., Ruckdeschel, G. Der Einfluss von Taurolin auf die körpereigene Abwehr und die Keimelimination bei der menschlichen Peritonitis Langenbecks. Arch Chir 1992; 377: 180-185.

63 Rosman, C., Westerveld, G.J., van Oeveren, W. et al. Effect of intraperitoneal antimicrobials on the concentration of bacteria, endotoxin, and tumor necrosis factor in abdominal fluid and plasma in rats Eur Surg Res 1996; 28: 351-360.

64 Traub, W.H., Leonhard, B., Bauer, D. Taurolidine: in vitro activity against multiple-antibiotic-resistant, nosocomially significant clinical isolates of Staphylococcus aureus, Enterococcus faecium, and diverse Enterobacteriaceae Chemother 1993; 39: 322-330.

65 Willatts, S.M., Radford, S., Leitermann, M. Effect of the antiendotoxic agent, taurolidine, in the treatment of sepsis syndrome: a placebo-controlled, double-blind trial Crit Care Med 1995; 23: 1033-1039.

66 Sherman, R.A. Maggot versus conservative debridement therapy for the treatment of pressure ulcers Wound Rep Regen 2002; 10: 208-214.

67 Sherman, R.A. Maggot therapy for treating diabetic foot ulcers unresponsive to conventional therapy. Diab Care 2003; 26: 446-451.

68 Sherman, R.A., Wyle, F., Vulpe, M. Maggot therapy for treating pressure ulcers in spinal cord injury patients. J Spinal Cord Med 1995; 18: 71-74.

69 Wayman, J., Nirojogi, V., Walker, A. et al. The cost effectiveness of larval therapy in venous ulcers. J Tissue Viabil 2000; 10: 91.

70 Courtenay, M., Whurch, J.C.T., Ryan, T.J. Larva

therapy in wound management. *J Royal Soc Med* 2000; 93: 72-74.

71 Daeschlein, G., Below, H., Hoffmeister, B. et al. Antibakterielle Effekte von Fliegenmaden (*Lucilia sericata*) in vitro zur Wundbehandlung *ZfW Sonderband* 2003.

72 Dissemond, J., Koppermann, M., Esser, S., et al. Therapie eines Methicillin-resistenten *Staphylokokkus aureus* (MRSA) im Rahmen der Behandlung eines chronischen Ulkus mittels *Biochirurgie Hautarzt* 2002; 53: 608-612.

73 Bonn, D. Maggot therapy: an alternative of wound infection. *Lancet* 2000; 356: 1174.

74 Grassberger, M. Wundbehandlung mit Fliegenlarven Teil 1 und Teil 2 *Hygiene Monitor Jahrgang* 2002; 8: 11/2002 und 12/2002

75 Gallenkemper, G. *Biochirurgie in der Behandlung von Problemwunden Teil 1 und 2 ZfW Nr. 1999 10/2 6-10 und 38-40*

76 Thomas, S., Andrews, A.M., Hay, N.P., Bourgoise, S. The anti-microbial activity of maggot secretions: results of a preliminary study *J Tiss Viabil* 1999; 9: 127-135.

77 Pavillard, E.R., Wright, E.A. An antibiotic from maggots. *Nature* 1957; 2: 916-917.

78 Prete, P.E. Growth effects of *phaenicia sericata* larval extracts on fibroblasts: mechanism for wound healing by maggot therapy *Life Sci* 1997; 60: 505-51097.

79 Hoekstra, M.J., Hupkens, P., Dutrieux, R.P. et al. A comparative burn wound model in the New Yorkshire pig for the histopathological evaluation of local therapeutic regimens: silver sulfadiazine cream as a standard *Br J Plast Surg* 1993; 46: 7, 585-589.

80 Zapata-Sirvent, R.L., Hansbrough, J.F. Cytotoxicity to human leukocytes by topical antimicrobial agents used for burn care. *Burn Care Rehabil* 1993; 14: 132-140.

81 McCauley, R.L., Li, Y.Y., Chopra, V. et al. Cytoprotection of human dermal fibroblasts against silver sulfadiazine using

recombinant growth factors *J Surg* 1994; 56: 378-384.

82 Kramer, A. Acriflavinumchlorid, Ethacridinlactat. In: Bruchhausen v.F, Ebel S, Fahm AW, Holzgrube U, Dannhardt G (Hrsg) *Hagers Handbuch der Pharmazeutischen Praxis, Stoffe, Springer, Berlin, Bd 7, 65, 1105. 1993*

83 Maitre, S., Jaber, K., Perrot, J.L. Increased serum and urinary levels of silver during treatment with topical silver sulfadiazine. *Ann Dermatol Venerol* 2002; 129: 217-219.

84 Goodman and Gilman's (1980): *The Pharmacological Basis of Therapeutics 6th Ed, MacMillan, New York, 977*

85 Bruck, J.C., Koch, S., Kramer, A. Klinische Untersuchungen zur Wirksamkeit von Lavasept auf granulierendem bzw. epithelisierenden Wunden *Hyg Med* 1998; 23: Supp 2:46

86 Trent, J.T, Kirsner, R.S. Necrotizing fasciitis. - *Wounds* 2002; 14: 8, 284-292.

87 Hingst, V., Vergetis, W. Antiseptische Sanierung von *Staphylococcus aureus*-Keimträgern in der Nase In: Kramer A, Gröschel D, Heeg P, Hingst V, Lippert H, Rotter M, Weuffen W (Hrsg) *Klinische Antiseptik. Springer, 1993.*

88 Kimura, M., Kawada, A. Contact sensitivity-induced by neomycin with cross-sensitivity to other aminoglycoside antibiotics *Contact Dermatitis* 1998; 39: 148-15015

89 Furr, J.R., Russel, A.D., Turner, T.D., Adrews, A. Antibacterial activity of Actisorb Plus, Actisorb and SILVER nitrate. *J Hosp Infect* 1994; 27: 201-208.

90 Müller, G., Winkler, Y., Kramer, A. Antibacterial activity and endotoxin-binding capacity of Actisorb Silver 220. *J Hosp Inf* 2003; 53: 211-214.

91 Lippert, H. (Hrsg), Kramer, A., Piatek, S. et al. (Mitarb) (2001): *Wundatlas: Wunde, Wundbehandlung und Wundheilung Barth, Heidelberg*

92 Wollina, U., Liebold, K., Schmidt, V.D. et al. Data and remittance spectroscopy measurement. *Int J Dermatol* 2002; 41: 635-639.

93 Fleischmann, W., Russ, M., Moch, D., Marquardt, C. *Biochirurgie – Sind Fliegenmaden wirklich die bessere. Chirurgie Chirurg* 1999; 70: 1340-1346.

94 Rudolph, P., Werner, H.P., Kramer, A. Untersuchungen zur Mikrobizidie von Wundaufflagen *Hyg Med* 2000; 25: 184-186.

95 Lyon, B.R., Skurray, R. Antimicrobial resistance of *staphylococcus aureus*: genetic basis. *Microbiol Rev* 1987; 51: 1:88-134.

96 Werner, H.P., Kramer, A. Mikrobiologische Anforderungen an lokale Antinfektiva unter spezieller Berücksichtigung der antiinfektiven Wundbehandlung In: Kramer A, Wendt M, Werner HP (Hrsg) *Möglichkeiten und Perspektiven der klinischen Antiseptik. mhp, Wiesbaden, 26-30 1995*

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<http://www.cdc.gov/niosh/rtecs/od481908.html>
www.octenisept.Schuelke-Mayr.de